


COLD STRESS: MOLECULAR EFFECTS ON MEDICINAL PLANTS

Selin Galatali^{1,a}

¹ *Mugla Sitki Kocman University, Faculty of Science, Molecular Biology and Genetics Department, Mugla, Turkey*

**Corresponding Author:*
E-mail: selingalatali@mu.edu.tr

(Received 24th March 2022; accepted 08th June 2022)

a:  ORCID 0000-0002-1992-3361

ABSTRACT. Plants do not have the ability to migrate to another suitable environment like other living things, for this reason, they should adapt to the different environmental conditions or their reproduction, the development and the growth are negatively affected. All these negative situations that affect plants negatively are called stress that are divided into two groups biotic and abiotic. The cold stress has two main effects on the plant: low temperature and dehydration. Plants create a tolerance response to this stress by laying out the transcriptional levels of proteins with different functions and especially transcription factors and some genes. Molecules involved directly or indirectly here include cryoprotectant proteins, chaperones, transcription factors and kinases. In the current review article, it was aimed to examine the cellular responses of medicinal plants treated with the cold stress during the adaptation process at the molecular level.

Keywords: *CAMTA, CBF/DREB TFs, cold stress, Hsp Family, MYB TF.*

INTRODUCTION

Medicinal plants have antimicrobial properties thanks to the secondary metabolites they contain, which allows them to be used for traditional medicine from ancient civilizations to the present. Along with the ever-developing scientific medicine, medicinal plants are still widely used around the world. The presence of approximately 20,000 medicinal plants, which are also the active ingredients of drugs used in the healing of many diseases, and for this reason, the World Health Organization (WHO) informed the countries for the use of traditional medicine in 1977 [1] and a year later, research and training centers were established in order to examine the medicinal plants in detail. As a result of the call for the establishment of medicinal plants, more use was encouraged [2]. Again, accordingly the research of the WHO [3], 80% of the developing countries and 60% of the countries around the world perform their health care by using traditional medicine [4, 5, 6].

Medicinal plants are widely used in a many of industries in the world. Due to the over side effects of artificial drugs produced in the pharmaceutical sector today, people use plants again to find healing in natural ways [7, 8]. In addition to taking an important place in the pharmaceutical industry with the active substances obtained from the secondary metabolites of medicinal plants, they are also used in cosmetics, spices, food, paints, insecticides, resins, gums, etc. It is also used in other areas, and its essential oils with aromatic properties are also used in the perfume industry [1, 9].

The secondary metabolites of plant, unlike primary metabolites, are not essential for the plant viability and metabolism, but are small molecule metabolism products necessary

for its secondary needs [10, 11]. Plants can synthesize secondary compounds including taxoids, polysaccharides, and flavones in addition to the basic nutrients they synthesize for their basic needs, such as protein, fat and carbohydrates, and in this way, the threats posed by bacteria, fungi and even other plants, as well as the biological stress they experience and environmental factors such as drought, temperature, humidity, UV rays and salinity. They can adapt to abiotic stresses caused by threats [12, 13]. At the same time, they communicate with other living things in which they are in harmony, and ensure the realization of events such as pollination and seed distribution, signaling, stimulating or inhibiting enzymes, acting as a cofactor for the catalytic activity of an enzyme [13, 14, 15]. Contrary to the sudden death of the plant in the absence of primary metabolites, in the absence or deficiency of secondary metabolites, adverse effects on the physical appearance of the plant, regression in reproduction and development are observed, while sometimes no change can be observed [16].

Plants are organisms that are in constant interaction with the external environment throughout their lives. As a result of changes in their environment, they do not have the ability to migrate to another suitable environment like other living things, and because they are sessile organisms, they either show the ability to adapt to the environmental condition changes or their reproduction, the development, the viability and the growth are negatively affected. All these negative situations that affect plants negatively are defined as stress [17, 18]. Some of these stresses are abiotic factors such as extreme cold, drought, excessive salinity, chemicals, UV radiation, while others are biotic factors created by microorganisms such as bacteria, fungi and predator animals [19]. Plant tolerance, however, refers to the continuation of the plant's potency to reproduce and survive by developing a physical, biochemical or molecular defense mechanism under these environmental and biological stress conditions [20]. In this review article, it was aimed to examine the cellular responses of medicinal plants treated with low temperature during the adaptation process at the molecular level. The present study is derived from a part of the related author's master's thesis.

The Abiotic Stress In Plants

Recently, as a result of climatic changes such as global warming, which tends to increase, and anthropogenic activities such as improper fertilization, sudden increases and decreases in temperatures, drought, excessive salinity, excess or insufficient amount of soil nutrients, changes in the amount of light and abiotic reactions such as UV rays. Stress factors, either alone or in combination, disrupt plant homeostasis, and this adversely affects plant growth, development and reproduction. Plants, like all other living things, suppress their developmental germination, reduce growth and reproduction against abiotic stresses in order to survive and continue their generation; physiologically, decrease in water intake, change in transmission rate, decrease in photosynthesis and nitrogen assimilation, change in respiration, accumulation of growth inhibitors; molecularly, it responds in the form of the gene expression regulation, synthesis of macromolecules, reduction of the activities of crucial enzymes, the synthesis reduction of protein and irregularity of membrane structure [21]. Depending on the severity and duration of any abiotic factors, the plant tolerance with its responses to this negative situation changes, and if the plant is subjected to excessive stress for a long time, it may not be able to tolerate this situation and result in death.

Among abiotic factors, cold stress is one of the most restrictive stresses for plant metabolism, development and growth. Cold stress has two main impacts on the plant: low temperature and dehydration. The cold effect occurs when the plant is exposed to temperatures below 15°C, which is called chilling temperature, or below 0°C, which is called freezing temperature (Fig. 1). At temperatures below 15°C, some limitations are observed in the enzyme activation included in the plant's respiration and photosynthesis processes [22, 23, 24] and biological events such as flowering, but the temperature level When it returns to an optimal level for the plant or when adaptation is in question, the plant continues its metabolic activities. However, at freezing temperatures under the 0°C, formation of ice begins in the extracellular area of plant cells and the water content in the soil, and this prevents water flow, causing dehydration and osmotic stress, and puts the plant on an irreversible path [21, 25, 26]. When dehydration occurs in the plant depending on the freezing temperature, ice formation begins in the intracellular area. This causes negative turgor pressure to occur and the protoplast volume to decrease. In addition, it can cause the formation of radicals that cause oxidative stress, denaturation of proteins, and changes in membrane potentials. As a result of intracellular and extracellular ice formation, it is the breakdown of plant tissues and membrane bilayer layer by freezing dehydration that will cause the worst damage to the plant [21, 25, 27, 28, 29].

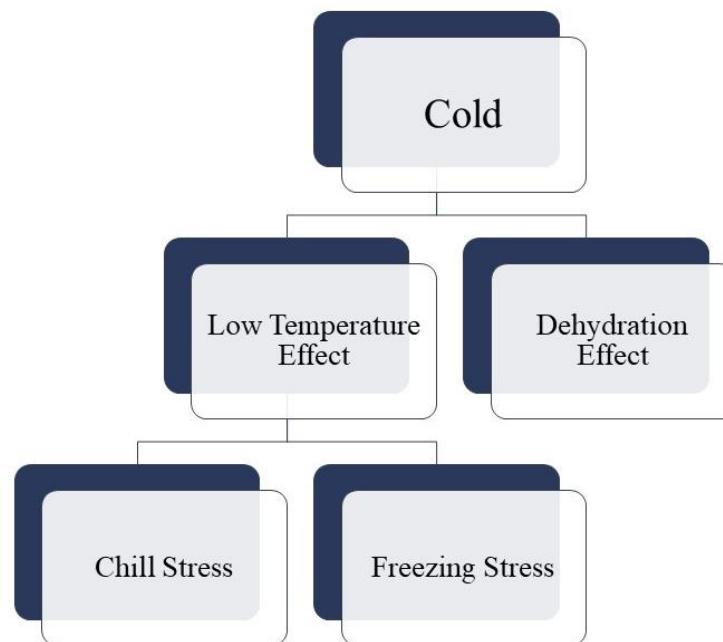


Fig. 1. Effects of cold stress on plants [25].

Different species show very different tolerances to cold temperatures to cope with cold stress. For example, plants that are sensitive to low temperature in tropical regions are damaged irreversibly as a result of deterioration of metabolism, alteration of protein and membrane structure, and inhibition of some enzymatic reactions even at cold temperatures, while plants that are tolerant to low temperatures and sensitive to freezing temperatures are susceptible to temperatures slightly below freezing. They can even survive, but they can get very risky damage as a result of their tissues freezing. In addition, plants that are tolerant to freezing temperatures can survive at these temperatures

depending on the cold treatment time and the freezing severity [30]. As a result, the extent of the damage that low temperatures will cause on the plant;

- the tissue type,
- the stage of growth and development,
- to freezing time
- and depending on the cooling rate.

When plants are faced to sudden variations in their environmental conditions, they try to adapt to the environment with some arrangements at the molecular level, despite the deterioration of their physiological and metabolic intracellular balances. When most plants are exposed to chilling temperatures, they increase the molecules accumulation such as osmolytes and cryoprotectants, which contain soluble sugars, and also change the cellular membrane composition. This process, called cold acclimation, helps plants tolerate frost stress [24, 31, 32]. When the plant experiences cold-induced stress at lower temperatures, it senses this by conformational changes in membrane stability and proteins as a result of the plasma membrane changing from a liquid crystal state to a solid gel state. This perception results in the transfer of the signal into the cell by causing the release of Ca^{2+} ions, which are the secondary messengers, from the cell surface via Ca^{2+} channels, and from within the cell from some organelles (mitochondria, endoplasmic reticulum) to the cytosol [33, 34, 35, 36, 37]. This triggers protein kinases and a number of transcription factor cascades, thereby activating various signal transduction pathways. Thus, the activation and/or inhibition of proteins such as cold-induced Hsp, LEA, COR, ICE is achieved, and eventually a molecular level response to stress occurs in the nucleus (Fig. 2).

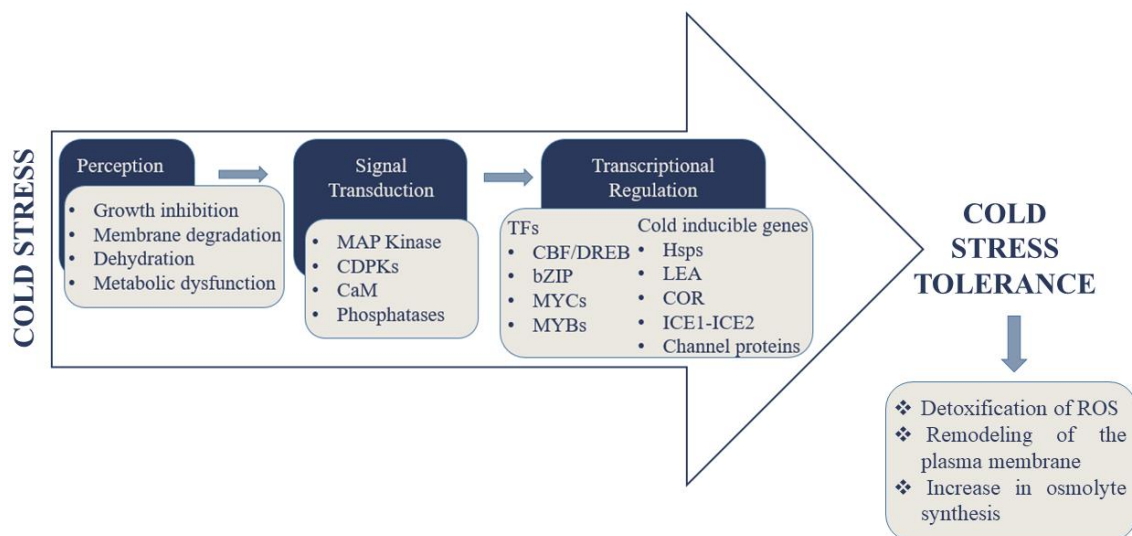


Fig. 2. A general molecular response to cold stress

Cold Stress Genes

Low temperatures are one of the most significant environmental stresses affecting plant productivity, growth, and development as well as their geographical distribution [69]. Plants create a tolerance response to this factor by arranging the transcriptional levels of proteins with different functions and especially transcription factors and some genes. Molecules involved directly or indirectly here include cryoprotectant proteins, chaperones, transcription factors and kinases [31, 37].

C-Repeat/Dehydration Responsive Element-Binding Factor

CBFs, also known as dehydration sensitive element binding factors (DREB), which are members of the AP2/ERF (APETALA2/Ethylene Sensitive Factor) transcription factor superfamily, play a role in the expression of cold-induced *COR* genes [24, 37, 38, 39]. For the protein expressions in charge of the cold stress response, CBF/DREB TFs bind to specific sequences (C-repeat: TGGCCCGAC) containing highly conserved CCGAC bases, also known as DRE/CRT, in the promoter site of cold and dehydration sensitive genes to positively control the expression of genes of interest [37, 40, 41, 42]. It has been reported by studies that proteins synthesized by CBF/DREB TFs in many plants are effective in the formation of cold tolerance by affecting the increase of the accumulation of proline, unsaturated fatty acids, and soluble sugars. There are some studies showing increased cold tolerance in some plant species of CBF TFs and in transgenic plants developed to overexpress *CBF/DREB* genes [43]. As seen in Table 1, CBF signal transduction pathway is very important for many plants in tolerating low temperature stress.

As shown in Fig. 4, transcription of *CBF* genes is regulated by some cold signaling molecules such as ICE, CAMTA, HOS, CCA, BZR and MYB [44, 45]. ICE, CAMTA, BZR and CCA regulate the positive gene expression by connecting to the promoter site of the *CBF* gene, while HOS1 negatively regulates the CBFs expression indirectly through 26S proteasome-mediated degradation of ICEs.

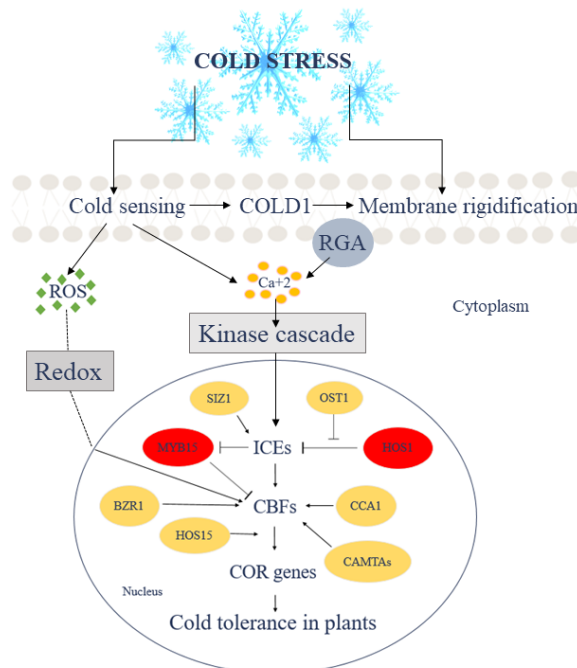


Fig. 3. The role of CBF transcription factors in the acquisition of cold stress tolerance [24].

MYB Transcription Factors

Plants manage to survive by arranging various signal networks with their wide range of TFs to adapt to the alteration of the environmental factors in their environment. MYBs are also one of the largest TFs families, accounting for approximately 9% of total TFs in plants. MYB proteins have MYB domains consisting of different repeats such as R1, R2,

R3, and R4 at their N-terminal ends and are basically divided into four subgroups according to these repeats. 1R-MYBs are classified as MYB-associated proteins, 2R-MYBs are R2R3-type MYB proteins, 3R-MYBs are R1R2R3-MYB proteins, and 4R-MYBs are classified as 4R-like MYB proteins [46, 47]. It has been stated in the works that *MYB* genes positively regulate the expression of many proteins included in the formation of the response to abiotic factors (Table 1). When studies on transgenic *A. thaliana* and *O. sativa* plants, which were developed to overexpress *MYB* genes, are examined, it is seen that *OsMYB3R-2* gene is effective in gaining tolerance against freezing, drought and salt stresses in *A. thaliana* [48]. It has provided tolerance to freezing and cold stress [49]. In *A. thaliana*, the *AtMYB15* gene was found to be effective in frost tolerance [50], and the *GmMYBJ1* gene in acquiring cold and drought tolerance [51], while the *OsMYB30* gene in *O. sativa* increased frost sensitivity [52] has been reported to be effective [37]. In another study, it was shown that PbrMYB5, an R2R3-type MYB gene in *Pyrus betulifolia*, positively modulated AsA synthesis, which is effective in cold stress tolerance, by binding to the promoter of the *PbrDCHAR2* gene as a transcriptional activator [47].

MYBs are TFs recognized to be effective in the cold stress response. MYBs also activate the relevant signal transduction pathways, as in other TFs, and also positively regulate the protein expression by connecting to the cis-elements located in the promoter site of target proteins (Fig. 4) [53].

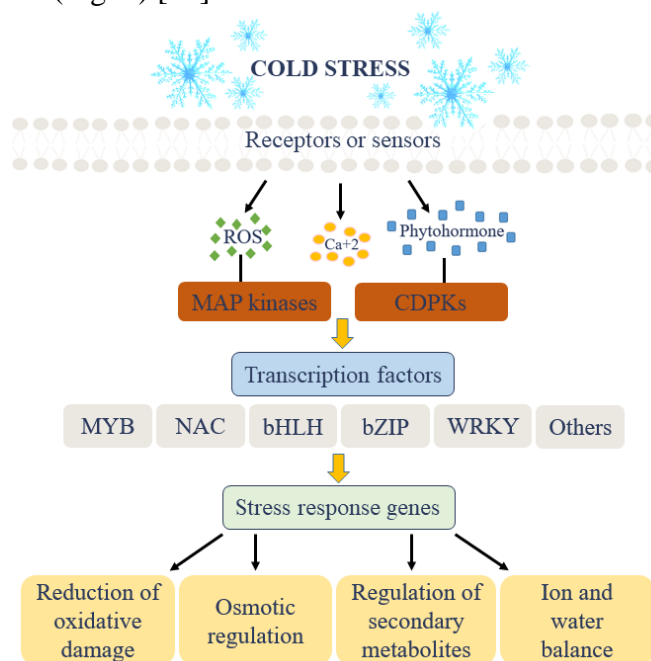


Fig. 4. The role of *myb* transcription factors in the acquisition of cold stress tolerance [53].

Calmodulin-Binding Transcription Activator

The calmodulin binding factors that contain the IQ domain for calmodulin binding belong to the CAMTA family. The N-terminal end of CAMTA proteins has the CG-1 domain, which shows properties of specific DNA binding. This domain binds to the vCGCGb sequence, also known as the CG-1 element, located 1 kb upstream from the start codon of the *CBF2* gene [45]. This interaction positively affects the expression of

the CBF2 transcription factor, which is known to play an active role in the cold stress response. As a result, the plant gains cold stress tolerance (Fig. 5).

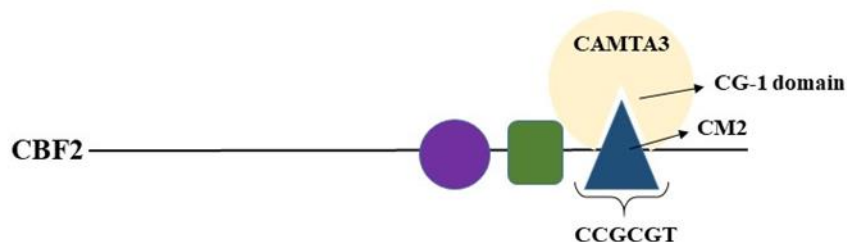


Fig. 5. The role of the CAMTA3 molecule in the acquisition of cold stress tolerance

Heat-Shock Protein Family

Abiotic stress factors such as cold and drought cause the proteins to deteriorate structurally and thus to lose their functions. Heat shock proteins, also known as molecular chaperones, are responsible for the membrane integrity and protein structures of plant cells in the absence of any stressor, thereby maintaining cellular homeostasis [54]. Under stress, the expression levels of Hsps are controlled by transcription factors called Hsfs. Hsp molecules, whose expression level changes with heat stress, contribute to the establishment of stress tolerance by performing events such as protein folding or degradation, and determination of protein localization [55]. For this reason, Hsps have a very important role against environmental stresses (water scarcity, heavy metals, cold, heat) experienced by plants. Among the heat shock proteins have been relatively well defined [54].

Table 1. Cold stress tolerance studies and observed effects on some natural and transgenic plants in the last ten years

Gene	Plant	Effect	Reference
<i>AnCBFs</i>	<i>Ammopiptanthus nanus</i>	Increased cold tolerance	[55]
<i>GthCBF4</i>	<i>Gossypium herbaceum</i>	Increased cold tolerance	[56]
<i>JcCBF2</i>	<i>Nicotiana benthamiana</i>	Enhanced drought tolerance	[57]
<i>VvCBF2</i> , <i>VvCBF3</i> , <i>VvCBF4</i> , <i>VvCBF6</i>	<i>Vitis vinifera</i> <i>V. riparia</i>	Cold tolerance	[58]
<i>AtDREB1A</i>	<i>Solanum lycopersicum</i>	Frost tolerance	[59]

<i>KcCBF3</i>	<i>Kandelia candel</i>	Cold tolerance	[60]
<i>MeCBF1</i>	<i>A. thaliana</i> <i>Manihot esculenta</i>	Increased cold tolerance	[61]
<i>GmDREB2A</i>	<i>Solanum melongena</i>	Increased cold tolerance	[62]
<i>PmhCBF_c</i>	<i>A. thaliana</i>	Increased freezing and oxidative stress tolerance	[63]
<i>GhCBF3</i>	<i>A. thaliana</i>	Increased drought and salt tolerance	[64]
<i>DaCBF7</i>	<i>O. sativa</i>	Enhanced cold tolerance	[65]
<i>GmDREB1</i>	<i>A. thaliana</i>	Cold, temperature and drought tolerance	[66]
<i>CbCBF</i>	<i>N. tabacum</i>	Frost tolerance	[67]
<i>NtDREB1A</i> <i>NtERD10B</i> <i>NtERD10C</i>	<i>Malus domestica</i> 'Gala'	Increased cold tolerance	[68]
<i>VaCBF1</i>	<i>N. tabacum</i>	Increased cold tolerance	[69]
<i>VaCBF4</i>	<i>A. thaliana</i>	Cold, salinity and drought tolerance	[70]
<i>ZmCBF3</i>	<i>O. sativa</i>	Cold, salinity and drought tolerance	[71]
<i>ZmDBP4</i>	<i>A. thaliana</i>	Enhanced cold and drought tolerance	[72]
<i>PpCBF1_v</i>	<i>Apple (Malus × domestica)</i>	Increased cold tolerance	[73]

<i>TaDREB2 TaDREB3</i>	<i>Triticum aestivum</i> <i>Hordeum vulgare</i>	Cold and drought tolerance	[74]
<i>CsR2R3-MYB</i>	<i>Camellia sinensis</i>	Increased cold tolerance	[75]
<i>GmMYB81</i>	<i>Glycine max</i>	Increased cold tolerance	[76]
<i>EgMY18,19,20</i>	<i>Elaeis guineensis</i>	Increased cold tolerance	[77]
<i>MdMYB108L</i>	<i>Malus communis</i>	Increased cold tolerance	[78]
<i>PbrMYB5</i>	<i>Pyrus betulaefolia</i>	Increased cold tolerance	[47]
<i>MdMYB124</i>	<i>Malus domestica</i>	Increased cold tolerance	[79]
<i>MdMYB88</i>	<i>A. thaliana</i>	Soğuşa dayanıklılığının artması	[79]
<i>OsMYBR1</i>	<i>O. sativa</i>	Increased cold tolerance	[80]
<i>FtMYB9</i>	<i>Fagopyrum tataricum</i>	Increased cold tolerance	[81]
<i>MYB96</i>	<i>A. thaliana</i>	Increased cold tolerance	[82]
<i>MdSIMYB1</i>	<i>M. domestica</i> 'Gala'	Increased cold tolerance	[68]
<i>TaMYB56-D</i>	<i>A. thaliana</i>	Increased cold tolerance	[83]
<i>TaMYB56-B</i>	<i>A. thaliana</i>	Increased cold tolerance	[83]
<i>R2R3 MYB</i>	<i>Cephalo f. tricolor</i>	Increased cold tolerance	[83]
<i>OsMYB2</i>	<i>O. Sativa</i>	Increased cold tolerance	[84]
<i>CAMTA</i>	<i>A. thaliana</i>	Increased cold tolerance	[85]

<i>TaCAMTA1-A, 1-D, 3-A ve 3-D</i>	<i>Triticum aestivum L.</i>	Increased cold tolerance	[86]
<i>CAMTA3 CAMTA5</i>	<i>A. thaliana</i>	Increased cold tolerance	[87]
<i>Camta3</i>	<i>A.thaliana</i>	Increased cold tolerance	[88]
<i>ZmCAMTA4a ZmCAMTA7a ZmCAMTA7b</i>	<i>Zea mays L.</i>	Increased cold tolerance	[89]
<i>Camta2</i>	<i>A.thaliana</i>	Increased cold tolerance	[90]
<i>Camta1</i>	<i>A.thaliana</i>	Increased cold tolerance	[90]
<i>CAMTA3</i>	<i>A. thaliana</i>	Increased cold tolerance	[45]
<i>HbHsfA4a HbHsfA4d, HbHsfA9b, HbHsfC1a HbHsfC1b</i>	<i>Hevea brasiliensis</i>	Increased cold tolerance	[91]
<i>RcHSP70</i>	<i>Rosa hybrida L.</i>	Increased cold tolerance	[92]
<i>QBS00812_HSP70</i>	<i>Lens culinaris</i>	Increased cold tolerance	[93]
<i>RcHSP70</i>	<i>R. hybrida L.</i>	Increased cold tolerance	[92]
<i>SlHSP17.7</i>	<i>S. lycopersicum L.</i>	Increased cold tolerance	[94]
<i>LpHSFC1</i>	<i>Lolium perenne L.</i>	Increased cold tolerance	[95]
<i>LpHSFC1b</i>	<i>L. perenne L.</i>	Increased cold tolerance	[95]
<i>HbHsfA4a</i>	<i>H. brasiliensis</i>	Increased cold tolerance	[96]
<i>HbHsfA1 HbHsfB1</i>	<i>H. brasiliensis</i>	Increased cold tolerance	[97]

KoHsp70 *G. hirsutum* Increased cold
tolerance [98]

CONCLUSIONS AND FUTURE PERSPECTIVES

Adaptation to the cold enables the low temperature resistant medicinal plants to gain frost resistance and to spend the winter months without being damaged by frost. The survival of these types of plants through the winter depends on their adaptability to the cold. Considering that the lowest average annual temperature is around 0 °C in most of the world and -10 °C in nearly half of it, the importance of cold adaptation and frost resistance on the yield to be obtained from agricultural activities in such environments will be better understood. Knowing the metabolic and molecular changes that occur in cold-resistant plant species and genotypes during cold adaptation will be beneficial both in terms of breeding studies and in terms of giving an idea about the species and genotype to be grown. Meeting the nutritional needs of the ever-increasing world population depends on the prevention of losses caused by low temperature stress. For this reason, breeding studies are carried out to obtain genotypes that are more tolerant to low temperatures, more adaptable to cold, and higher frost resistance.

REFERENCES

- [1] Faydaoglu, E., Surucuoglu, M.S. (2011): The use and economic importance of medicinal and aromatic plants from past to present. *Kastamonu University Journal of Forestry Faculty* 11 (1): 52-67.
- [2] Acibuca, V., Budak, D.B. (2018): The place and importance of medicinal and aromatic plants in the world and in Turkey. *Çukurova Journal of Agricultural and Food Sciences* 33 (1): 37-44.
- [3] WHO (1993): <http://apps.who.int/iris/bitstream/handle/10665/36838/9241544465.pdf;jsessionid=8081FBAC05BB1C37AFCAA9377A69CA41?sequence=1>
- [4] Ozudogru, E.A., Kaya E, Kirdok, E., Issever-Ozturk, S. (2011a): *In vitro* propagation from young and mature explants of thyme (*Thymus vulgaris* and *T. longicaulis*) resulting in genetically stable shoots. *In Vitro Cellular & Developmental Biology – Plant* 47(2): 309-320.
- [5] Ozudogru, E.A., Kaya, E., Kirdok, E. (2011b): Development of protocols for short-, medium- and long-term conservation of thyme. *Acta Horticulturae* 918: 43-50.
- [6] Mahajan, M., Kuiry, R., Pal, P.K. (2020): Understanding the consequence of environmental stress for accumulation of secondary metabolites in medicinal and aromatic plants. *Journal of Applied Research on Medicinal and Aromatic Plants* 18: 100255.
- [7] Munda, S., Dutta, S., Pandey, S.K., Sarma, N., Lal, M. (2019): Antimicrobial activity of essential oils of medicinal and aromatic plants of the North East India: A biodiversity hot spot. *Journal of Essential Oil-Bearing Plants* 22 (1): 105-119.
- [8] Kaya, E., Balci, M.A., Akguller, O., Galatali, S., Yeniocak, S., Mercan, T., Guldag, S., Ozkaya, D.E., Ozturk, B., Celik, O., Aktay, I. (2021): Development of an optimum proliferation medium via the graph kernel statistical analysis method for genetically stable *in vitro* propagation of endemic *Thymus cilicicus* (Turkey). *Acta Botanica Croatica* 80(2): 199-207.
- [9] Kaya, E., Alves, A., Rodrigues, L., Jenderek, M., Hernandez-Ellis, M., Ozudogru, E.A., Ellis, D. (2013): Cryopreservation of Eucalyptus Genetic Resources. *Cryoletters* 34(6): 608-618.

- [10] Yang, L., Wen, K.S., Ruan, X., Zhao, Y.X., Wei, F., Wang Q. (2018): Response of plant secondary metabolites to environmental factors. *Molecules* 23 (4): 762.
- [11] Kivrak, S., Gokturk, T., Kivrak, I., Kaya, E., Karababa, E. (2019): Investigation of phenolic profiles and antioxidant activities of some *Salvia* species commonly grown in Southwest Anatolia using UPLC-ESI-MS/MS. *Food Science and Technology* 32(2): 423-431.
- [12] Bennett, R.N., Wallsgrave, R.M. (1994): Secondary metabolites in plant defence mechanisms. *New Phytologist* 127 (4): 617-633.
- [13] Wink, M. (2008): Evolution of secondary plant metabolism. eLS.
- [14] Wink, M. (2003): Evolution of secondary metabolites from an ecological and molecular phylogenetic perspective, *Phytochemistry* 64 (1): 3-19.
- [15] Zhao, J., Davis, L.C., Verpoorte, R. (2005): Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnology Advances* 23 (4): 283-333.
- [16] Tiwari, R., Rana, C. (2015): Plant secondary metabolites: a review. *International Journal of Engineering* 3 (5): 661-670.
- [17] Buyuk, I., Soydam Aydin, S., Sumer, A. (2012): Molecular responses of plants to stress conditions. *Turkish Journal of Hygiene and Experimental Biology* 69 (2): 97-110.
- [18] Ozudogru, E.A., Kaya, E. (2012): Cryopreservation of *Thymus cariensis* and *T. vulgaris* shoot tips comparison of there vitrification based methods. *Cryoletters* 33(5): 363-375.
- [19] Lichtenthaler, H.K. (1996): Vegetation stress: an introduction to the stress concept in plants. *Journal of Plant Physiology* 148 (1-2): 4-14.
- [20] Simms, E.L. (2000): Defining tolerance as a norm of reaction. *Evolutionary Ecology* 14 (4): 563-570.
- [21] Donmez, C. (2018): Determination of mRNA expression levels of bZIP, MYB, NAC, NF-YB and WRKY type transcription factor genes in sunflower (*Helianthus annuus* L.) plant grown under cold stress, Integrated PhD Thesis, Ankara University, Ankara, Turkey, 252s.
- [22] O'Kane, D., Gill, V., Boyd, P., Burdon, R. (1996): Chilling, oxidative stress and antioxidant responses in *Arabidopsis thaliana* callus. *Planta* 198 (3): 371-377.
- [23] Yang, M.T., Chen, S.L., Lin, C.Y., Chen, Y.M. (2005): Chilling stress suppresses chloroplast development and nuclear gene expression in leaves of mung bean seedlings. *Planta* 221 (3): 374-385.
- [24] Shi, Y., Yang, S. (2014): ABA regulation of the cold stress response in plants. *Abscisic acid: metabolism, transport and signaling*. Springer. Dordrecht, pp. 337-363.
- [25] Beck, E.H., Heim, R., Hansen, J. (2004): Plant resistance to cold stress: mechanisms and environmental signals triggering frost hardening and dehardening. *Journal of Biosciences* 29 (4): 449-459.
- [26] Lindlöf, A., Chawade, A., Sikora, P., Olsson, O. (2015): Comparative transcriptomics of Sijung and Jumli Marshi rice during early chilling stress imply multiple protective mechanisms. *PLOS ONE* 10 (5): e0125385.
- [27] Steponkus, P.L. (1984): Role of the plasma membrane in freezing injury and cold acclimation. *Annual Review of Plant Physiology* 35 (1): 543-584.
- [28] Steponkus, P. (1993): A contrast of the cryostability of the plasma membrane of winter rye and spring oat-two species that widely differ in their freezing tolerance and plasma membrane lipid composition. *Advances in Low-Temperature Biology* 3: 211-312.
- [29] Ozkaya, D.E., Celik, O., Galatali, S., Kaya, E. (2021): Investigation of Cold Stress Effects on Plants at Molecular Level. *SunText Review of BioTechnology* 2(2): 1-7.
- [30] Hirt, H., Shinozaki, K. (2003): Plant responses to abiotic stress. *SSBM*.
- [31] Thomashow, M.F. (1999): Plant cold acclimation: freezing tolerance genes and regulatory mechanisms. *Annual Review of Plant Physiology* 50 (1): 571-599.
- [32] Yamada, T., Kuroda, K., Jitsuyama, Y., Takezawa, D., Arakawa, K., Fujikawa, S. (2002): Roles of the plasma membrane and the cell wall in the responses of plant cells to freezing. *Planta* 215 (5): 770-778.
- [33] Yan, Y., Wei, C. L., Zhang, W. R., Cheng, H. P., Liu, J. (2006): Cross-talk between calcium and reactive oxygen species signaling. *Acta Pharmacologica Sinica* 27(7): 821-826.

- [34] Ruelland, E., Vaultier, M.N., Zachowski, A., Hurry, V. (2009): Chapter 2 cold Signalling and cold acclimation in plants. *Advances in Botanical Research* 49: 35-150.
- [35] Ruelland, E., Zachowski, A. (2010): How plants sense temperature. *Environmental and Experimental Botany* 69(3): 225-232.
- [36] Chinnusamy, V., Zhu, J.K., Sunkar, R. (2010): Gene regulation during cold stress acclimation in plants. In: Sunkar, R. (ed.) *Plant Stress Tolerance*. Stillwater, ABD, pp. 39-55.
- [37] Mehrotra, S., Verma, S., Kumar, S., Kumari, S., Mishra, B.N. (2020): Transcriptional regulation and signalling of cold stress response in plants: an overview of current understanding. *Environmental and Experimental Botany* 104243.
- [38] Liu, Q., Kasuga, M., Sakuma, Y., Abe, H., Miura, S., Yamaguchi-Shinozaki, K., Shinozaki, K. (1998): Two transcription factors, DREB1 and DREB2, with an EREBP/AP2 DNA binding domain separate two cellular signal transduction pathways in drought-and low-temperature-responsive gene expression, respectively, in *Arabidopsis*. *The Plant Cell* 10 (8): 1391-1406.
- [39] Zhou, M.L., Ma, J.T., Pang, J.F, Zhang, Z.L., Tang, Y.X., Wu, Y.M. (2010): Regulation of plant stress response by dehydration responsive element binding (DREB) transcription factors. *African Journal of Biotechnology* 9 (54): 9255-9269.
- [40] Chew, Y.H., Halliday, K.J. (2011): A stress-free walk from *Arabidopsis* to crops. *Current Opinion Journals* 22 (2): 281-286.
- [41] Rihan, H.Z., Al-Issawi, M., Fuller, M.P. (2017): Advances in physiological and molecular aspects of plant cold tolerance. *Journal of Plant Interactions* 12 (1): 143-157.
- [42] Vazquez-Hernandez, Romero, M., I., Escribano, M.I., Merodio, C., Sanchez-Ballesta, M.T. (2017): Deciphering the role of CBF/DREB transcription factors and dehydrins in maintaining the quality of table *grapes* cv. autumn royal treated with high CO₂ levels and stored at 0 C. *Frontiers in Plant Science* 8: 1591.
- [43] Ito, Y., Katsura, K., Maruyama, K., Taji, T., Kobayashi, M., Seki, M., Yamaguchi-Shinozaki K. (2006): Functional analysis of rice DREB1/CBF type transcription factors involved in cold responsive gene expression in transgenic rice. *Plant and Cell Physiology* 47 (1): 141-153.
- [44] Dong, C.H., Agarwal, M., Zhang, Y., Xie, Q., Zhu, J.K. (2006): The negative regulator of plant cold responses, HOS1, is a RING E3 ligase that mediates the ubiquitination and degradation of ICE1. *Proceedings of the National Academy of Sciences of the United* 103 (21): 8281-8286.
- [45] Doherty, C.J., Buskirk, H.A.V., Myers, S.J., Thomashow, M.F. (2009): Roles for *Arabidopsis* CAMTA transcription factors in cold-regulated gene expression and freezing tolerance. *The Plant Cell* 21 (3): 972-984.
- [46] Lipsick, J.S. (1996): One billion years of MYB. *Oncogene* 13 (2): 223-235.
- [47] Xing, C., Liu, Y., Zhao, L., Zhang, S., Huang, X. (2019): A novel MYB transcription factor regulates ascorbic acid synthesis and affects cold tolerance. *Plant, Cell & Environment* 42(3): 832-845.
- [48] Dai, X., Xu, Y., Ma, Q., Xu, W., Wang, T., Xue, Y., Chong, K. (2007): Overexpression of an R1R2R3 MYB gene, OsMYB3R-2, increases tolerance to freezing, drought, and salt stress in transgenic *Arabidopsis*. *Plant Physiology* 143 (4): 1739-1751.
- [49] Ma, Q., Dai, X, Xu, Y., Guo, J., Liu, Y., Chen, N., Zhang, X. (2009): Enhanced tolerance to chilling stress in OsMYB3R-2 transgenic rice is mediated by alteration in cell cycle and ectopic expression of stress genes. *Plant Physiology* 150 (1): 244-256.
- [50] Agarwal, M., Hao, Y., Kapoor, A., Dong, C.H., Fujii, H., Zheng, X., Zhu, J.K. (2006): A R2R3 type MYB transcription factor is involved in the cold regulation of CBF genes and in acquired freezing tolerance. *International Journal of Biological Chemistry* 281: 37636–37645.

- [51] Su, L.T., Li, J.V., Liu, D.Q., Zhai, Y., Zhang, H.J., Li, X.W., Zhang, Q.L., Wang, Y., Wang, Q.Y. (2014): A novel MYB transcription factor, GmMYBJ1, from soybean confers drought and cold tolerance in *Arabidopsis thaliana*. *Gene* 538 (1): 46-55.
- [52] Lv, Y., Yang, M., Hu, D., Yang, Z., Ma, S., Li, X., Xiong, L. (2017): The OsMYB30 transcription factor suppresses cold tolerance by interacting with a JAZ protein and suppressing β -amylase expression. *Plant Physiology* 173 (2): 1475-1491.
- [53] Khan, S.A., Li, M.Z., Wang, S.M., Yin, H.J. (2018): Revisiting the role of plant transcription factors in the battle against abiotic stress. *International Journal of Molecular Sciences* 19 (6): 1634.
- [54] Krishna, P., Sacco, M., Cherutti, J.F., Hill, S. (1995): Cold induced accumulation of Hsp90 transcripts in *Brassica napus*. *Plant Physiology* 107 (3): 915-923.
- [55] Cheng, Y. Bai, S., Zhang, L.P., Chen, L., Fan, G., Xu, J.G., Guo, Z.F. (2021): Identification and characterization of AnICE1 and AnCBFs involved in cold tolerance from *Ammopiptanthus nanus*. *Plant Physiology and Biochemistry* 168: 70-82.
- [56] Liu, J., Magwanga, R., Xu, Y., Wei, T., Zheng, J., Hou, Y., Liu, F. (2021): Functional characterization of cotton c-repeat binding factor genes reveal their potential role in cold stress tolerance. *Frontiers in Plant Science* 12, 766130-766130.
- [57] Wang, L., Wu, Y., Tian, Y., Dai, T., Xie, G., Xu, Y., Chen, F. (2020): Overexpressing *Jatropha curcas* CBF2 in *Nicotiana benthamiana* improved plant tolerance to drought stress. *Gene* 742: 144588.
- [58] Rubio, S., Noriega, X., Pérez, F.J. (2019): Abscisic acid (ABA) and low temperatures synergistically increase the expression of CBF/DREB1 transcription factors and cold-hardiness in grapevine dormant buds. *Annals of Botany* 123(4): 681-689.
- [59] Karkute, S. G., Krishna, R., Ansari, W. A., Singh, B., Singh, P. M., Singh, M., Singh, A. K. (2019): Heterologous expression of the AtDREB1A gene in tomato confers tolerance to chilling stress. *Biologia Plantarum* 63(1): 268-277.
- [60] Du, Z., Li, J. (2019): Expression, purification and molecular characterization of a novel transcription factor KcCBF3 from *Kandelia candel*. *Protein Expression and Purification* 153: 26-34.
- [61] An, D., Ma, Q., Wang, H., Yang, J., Zhou, W., Zhang, P. (2017): Cassava C-repeat binding factor 1 gene responds to low temperature and enhances cold tolerance when overexpressed in *Arabidopsis* and cassava. *Plant molecular biology* 94(1): 109-124.
- [62] Yamasaki, Y., Randall, S.K. (2016): Functionality of soybean CBF/DREB1 transcription factors. *Plant Science* 246: 80-90.
- [63] Peng, T., Guo, C., Yang, J., Xu, M., Zuo, J., Bao, M., Zhang, J. (2016): Overexpression of a Mei (*Prunus mume*) CBF gene confers tolerance to freezing and oxidative stress in *Arabidopsis*. *Plant Cell, Tissue and Organ Culture* 126(3): 373-385.
- [64] Ma, L. F., Li, Y., Chen, Y., Li, X. B. (2016): Improved drought and salt tolerance of *Arabidopsis thaliana* by ectopic expression of a cotton (*Gossypium hirsutum*) CBF gene. *Plant Cell, Tissue and Organ Culture* 124(3): 583-598.
- [65] Byun, M. Y., Lee, J., Cui, L. H., Kang, Y., Oh, T. K., Park, H., Lee, H., Kim, W. T. (2015): Constitutive expression of DaCBF7, an Antarctic vascular plant *Deschampsia antarctica* CBF homolog, resulted in improved cold tolerance in transgenic rice plants. *Plant Science* 236: 61-74.
- [66] Kidokoro, S., Watanabe, K., Otori, T., Moriwaki, T., Maruyama, K., Mizoi, J., Yamaguchi-Shinozaki, K. (2015): Soybean DREB 1/CBF-type transcription factors function in heat and drought as well as cold stress-responsive gene expression. *The Plant Journal* 81(3): 505-518.
- [67] Zhou, M., Xu, M., Wu, L., Shen, C., Ma, H., Lin, J. (2014): CbCBF from *Capsella bursa-pastoris* enhances cold tolerance and restrains growth in *Nicotiana tabacum* by antagonizing with gibberellin and affecting cell cycle signaling. *Plant Molecular Biology* 85(3): 259-275.

- [68] Wang, R. K., Cao, Z. H., Hao, Y. J. (2014): Overexpression of a R2R3 MYB gene MdSIMYB1 increases tolerance to multiple stresses in transgenic tobacco and apples. *Physiologia Plantarum* 150(1): 76-87.
- [69] Dong, C., Zhang, M., Yu, Z., Ren, J., Qin, Y., Wang, B., Tao, J. (2013): Isolation and expression analysis of CBF4 from *Vitis amurensis* associated with stress. *The Journal of Agricultural Science*.
- [70] Li, Y., Yu, W., Lu, Q., Yang, S., Tian, W. (2013): Effects of abiotic stresses on the expression of Heat Shock Transcription Factor (HSF) family members in *Rubber tree* (*Hevea brasiliensis* Muell. Arg.). *CJTC* 42(8): 2119.
- [71] Xu, M., Li, L., Fan, Y., Wan, J., Wang, L. (2011): ZmCBF3 overexpression improves tolerance to abiotic stress in transgenic rice (*Oryza sativa*) without yield penalty. *Plant cell reports* 30(10): 1949-1957.
- [72] Wang, C. T., Yang, Q., Yang, Y. M. (2011): Characterization of the ZmDBP4 gene encoding a CRT/DRE-binding protein responsive to drought and cold stress in maize. *Acta Physiologiae Plantarum* 33(2): 575-583.
- [73] Wisniewski, M., Norelli, J., Bassett, C., Artlip, T., Macarisin, D. (2011): Ectopic expression of a novel peach (*Prunus persica*) CBF transcription factor in apple (*Malus domestica*) results in short-day induced dormancy and increased cold hardiness. *Planta* 233(5): 971-983.
- [74] Morran, S., Eini, O., Pyvovarenko, T., Parent, B., Singh, R., Ismagul, A., Eliby, S., Shirley, N., Langridge, P., Lopato, S. (2011): Improvement of stress tolerance of wheat and barley by modulation of expression of DREB/CBF factors. *Plant Biotechnology Journal* 9(2): 230-249.
- [75] Chen, X., Wang, P., Gu, M., Lin, X., Hou, B., Zheng, Y., Ye, N. (2021): R2R3-MYB transcription factor family in tea plant (*Camellia sinensis*): Genome-wide characterization, phylogeny, chromosome location, structure and expression patterns. *Genomics* 113(3): 1565-1578.
- [76] Bian, S., Jin, D., Sun, G., Shan, B., Zhou, H., Wang, J., Li, X. (2020): Characterization of the soybean R2R3-MYB transcription factor GmMYB81 and its functional roles under abiotic stresses. *Gene* 753: 144803.
- [77] Zhou, L., Yarra, R., Jin, L., Cao, H. (2020): Genome-wide identification and expression analysis of MYB gene family in oil palm (*Elaeis guineensis* Jacq.) under abiotic stress conditions. *Environmental and Experimental Botany* 180: 104245.
- [78] Wang, Y., Mao, Z., Jiang, H., Zhang, Z., Chen, X. (2019): A feedback loop involving MdMYB108L and MdHY5 controls apple cold tolerance. *Biochemical and Biophysical Research Communications* 512(2): 381-386.
- [79] Xie, Y., Chen, P., Yan, Y., Bao, C., Li, X., Wang, L., Shen, X., Li, H., Liu, X., Niu, C., Zhu, C., Fang, N., Shao, Y., Zhao, T., Yu, J., Zhu, J., Xu, L., van Nocker, S., Ma, F., Guan, Q. (2018): An atypical R2R3 MYB transcription factor increases cold hardiness by CBF-dependent and CBF-independent pathways in apple. *New Phytologist* 218(1): 201-218.
- [80] Yin, X., Cui, Y., Wang, M., Xia, X. (2017): Overexpression of a novel MYB-related transcription factor, OsMYBR1, confers improved drought tolerance and decreased ABA sensitivity in rice. *Biochemical and Biophysical Research Communications* 490(4): 1355-1361.
- [81] Gao, F., Zhou, J., Deng, R.Y., Zhao, H.X., Li, C.L., Chen, H., Wu, Q. (2017): Overexpression of a *Tartary buckwheat* R2R3-MYB transcription factor gene, FtMYB9, enhances tolerance to drought and salt stresses in transgenic Arabidopsis. *Journal of Plant Physiology* 214: 81-90.
- [82] Lee, S. B., Suh, M. C. (2015): Advances in the understanding of cuticular waxes in Arabidopsis thaliana and crop species. *Plant Cell Reports* 34(4): 557-572.
- [83] Zhang, B., Hu, Z., Zhang, Y., Li, Y., Zhou, S., Chen, G. (2012): A putative functional MYB transcription factor induced by low temperature regulates anthocyanin biosynthesis in

- purple kale (*Brassica oleracea* var. *acephala* f. *tricolor*). *Plant Cell Reports* 31(2): 281-289.
- [84] Yang, A., Dai, X., Zhang, W. H. (2012): A R2R3-type MYB gene, OsMYB2, is involved in salt, cold, and dehydration tolerance in rice. *Journal of Experimental Botany* 63(7): 2541-2556.
- [85] Xiao, P., Feng, J.W., Zhu, X.T., Gao, J. (2021): Evolution analyses of CAMTA transcription factor in plants and its enhancing effect on cold-tolerance. *Frontiers in Plant Science* 12.
- [86] Tang, T., Li, C.H., Li, D.S., Jing, S.X., Hua, J., Luo, S.H., Li, S.H. (2020): Peltate glandular trichomes of *Colquhounia vestita* harbor diterpenoid acids that contribute to plant adaptation to UV radiation and cold stresses. *Phytochemistry* 172: 112285.
- [87] Kidokoro, S., Yoneda, K., Takasaki, H., Takahashi, F., Shinozaki, K., Yamaguchi-Shinozaki, K. (2017): Different cold signaling pathways function in the responses to rapid and gradual decreases in Temperature. *The Plant Cell* 29(4): 760-774.
- [88] Kim, Y.S., An, C., Park, S., Gilmour, S.J., Wang, L., Renna, L., Thomashow, M.F. (2017): CAMTA mediated regulation of salicylic acid immunity pathway genes in *Arabidopsis* exposed to low temperature and pathogen infection. *The Plant Cell* 29(10): 2465-2477.
- [89] Yue, R., Lu, C., Sun, T., Peng, T., Han, X., Qi, J., Yan, S., Tie, S. (2015): Identification and expression profiling analysis of calmodulin-binding transcription activator genes in maize (*Zea mays* L.) under abiotic and biotic stresses. *Frontiers in Plant Science* 6: 576.
- [90] Kim, Y., Park, S., Gilmour, S.J., Thomashow, M.F. (2013): Roles of CAMTA transcription factors and salicylic acid in configuring the low-temperature transcriptome and freezing tolerance of *Arabidopsis*. *The Plant Journal* 75(3): 364-376.
- [91] Yan, L., Wencai, Y., Qingzhi, L., Shuguang, Y., Weimin, T. (2021): Effects of Abiotic Stresses on the Expression of Heat Shock Transcription Factor (HSF) Family Members in Rubber Tree (*Hevea brasiliensis* Muell. Arg.). *Chinese Journal of Tropical Crops* 42(8): 2119.
- [92] Jiang, C., Bi, Y., Zhang, R., Feng, S. (2020): Expression of RcHSP70, heat shock protein 70 gene from *Chinese rose*, enhances host resistance to abiotic stresses. *Scientific Reports* 10(1): 1-10.
- [93] Khorshidvand, M., Ismaili, A., Sohrabi, S.S., Haghjou, M.M. (2021): Identification and expression pattern of lentil's HSPs under different abiotic stresses. *Plant Biotechnology Reports* 15(5): 609-625.
- [94] Zhang, N., Zhao, H., Shi, J., Wu, Y., Jiang, J. (2020): Functional characterization of class I SIHSP17.7 gene responsible for tomato cold-stress tolerance. *Plant Science* 298: 110568.
- [95] Sun, T., Shao, K., Huang, Y., Lei, Y., Tan, L., Chan, Z. (2020): Natural variation analysis of *Perennial ryegrass* in response to abiotic stress highlights LpHSFC1b as a positive regulator of heat stress. *Environmental and Experimental Botany* 179: 104192.
- [96] Wencai, Y., Yan, L., Shaohua, W., Jinquan, C., Shixin, Z., Shuguang, Y., Weimin, T. (2019): Cloning of HbHsfA4a and Its Transcriptional Responses to Cold Stress in *Hevea brasiliensis*. *Chinese Journal of Tropical Crops* 40(5): 898.
- [97] Deng, X., Wang, J., Wang, J., Tian, W. (2018): Two HbHsfA1 and HbHsfB1 genes from the tropical woody plant rubber tree confer cold stress tolerance in *Saccharomyces cerevisiae*. *Revista Brasileira de Botanica* 41(3): 711-724.
- [98] Fei, J., Wang, Y.S., Zhou, Q., Gu, J.D. (2015): Cloning and expression analysis of HSP70 gene from mangrove plant *Kandelia obovata* under cold stress. *Ecotoxicology* 24(7): 1677-1685.